

天然物化学における単離について

—海洋天然物化学 特に核酸 テトラトキノン及びピテトラトキノン類化合物の分離について—
韓国科学院 金容海

Isolation, Purification and Structural study of Marine Natural
Products of Nucleic Acids, Tetrodotoxin and Chiriquitoxin

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Summary Separation and structural study of biologically active marine natural products are discussed. A rare nucleic acid, doridosine, was isolated from boneless sea animals, dorid nudibranch, *Anisodoris nobilis*, digestive glands shown to cause prolonged reduced arterial pressure and heart rate in mammals. The structure of doridosine was determined as N¹-methyl isoguanosine. Potent neurotoxins of tetrodotoxin (TTX) and chiriquitoxin (CHTX) were isolated from *Atelopus chiriquiensis* frogs. CHTX shows similar biological activity and has 392 molecular weight (73 higher than TTX (319)). Biologically active (anti-histamine release effect) material was isolated from *Ligia exotica* together with various nucleic acids and amino acids.

1 Introduction

A new rare nucleic acid, N-methylnucleoside named doridosine, identified as 1-methylisoguanosine, was isolated from the shell-less marine dorid nudibranch *Anisodoris nobilis*, from the Australian sponge, *Tadania digitata*^{2,3} recently from the Caribbean coral *Madaraca mirabilis*⁴. While, isoguanosine was isolated from dorid nudibranch *Diaulula sandigensis*⁵. Recently, inosine and unknown nucleic acid were isolated and confirmed from *Ligia exotica*.

Based on the hypothesis that shell-less marine mollusks, specifically dorid nudibranchs, might contain a toxic substance that protects them from predators, various tissue extracts of specimens were prepared⁶. Aqueous extracts of these were subjected to a preliminary purification by dialysis followed by general toxicity by injection into mice and crabs. Toxic extracts were studied in more detail in several standard pharmacological preparations. From these studies it emerged that extracts of the digestive glands of *Anisodoris nobilis* had the unusual property,

among extracts from marine animals, of producing hypotension and bradycardia in mammals within a few seconds of intravenous injection in anesthetized rats. These extracts caused a marked and prolonged reduction in heart rate and a sharp drop in systolic blood pressure. The activity of the extracts could be followed conveniently by testing on the isolated, spontaneously beating, guinea pig atria. We reported that the cardioactive component of the digestive gland of Anisodoris nobilis is a new N-methylpurine riboside that we named doridosine. Doridosine was assigned the structure of 1-methylisoguanosine (1) based on its spectroscopic properties. The function and origin of this compound in these marine organisms are unknown. It is known that dorid nudibranchs feed principally on sponges,^{7,8} but our preliminary studies have failed to detect any doridosine in several food sources of Anisodoris nobilis.

Doridosine causes reduced arterial pressure and reduced heart rate in mammals in a manner that is qualitatively similar to adenosine but with an unusually long duration of action.^{9,10} It also shows skeletal muscle relaxant and hypothermic activity.^{9,10}

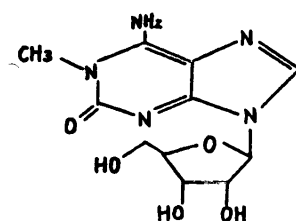
It is interesting that various nucleic acids of inosine, uracil, guanosine and 2-deoxyguanosine, and anti-allergy compound were isolated from Ligia exotica. So small sea animal contains large amounts of nucleic acids and various amino acids. Low pressure liquid column chromatography (Bio-Gel P2 and Sephadex G10) and JAI (Japan Analytical Institute) HPLC (column GAIGS 320 20 x 500 mm) combination was very effective for the separation of various amino acids and nucleic acids.

Chiriquitoxin (CHTX, 1) was first isolated in 1975 from the Costa Rican frog Atelopus chiriquiensis.¹¹ On the basis of ¹H NMR¹² and mass spectra¹³, its structure was postulated to differ from that of tetrodotoxin (TTX, 2) only with

respect to the substituent at C-6 2 is a potent neurotoxin of puffers¹⁴ and newts,¹⁵ and is an important neurobiological tool¹⁶ Among derivatives and natural analogs of 2,¹⁷ 1 is unique in being as potent as 2 in lethality to mice¹¹ and in blocking the voltage-gated sodium channel,¹⁸ whereas all others have markedly reduced biological activities¹⁹ Earlier work with 1 was hampered by a scarcity of material and by difficulties in separating it from co-existing 2 In late June 1988, renewed collection of the frogs was successful Using the paradigm which led to structural determination of natural analogs of 2 in newts^{17a} and in puffers^{17b,c,d} We report here the structure of 1

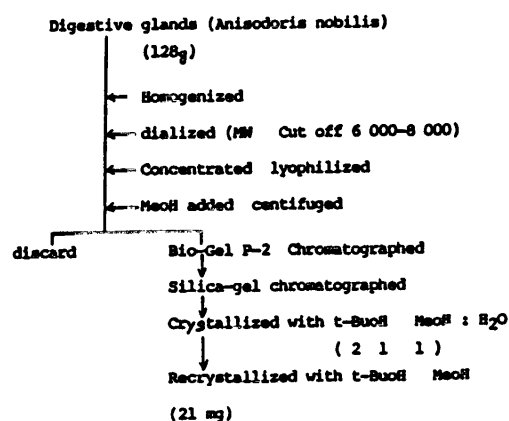
2 Doridosine (1-methyl isoguanosine)

This marine natural product was isolated from dorid nudibranch, Anisodoris nobilis (digestive glands) As a rare nucleic acids of isoguanosine derivatives, its structure was elucidated as 1-methylisoguanosine Separation and purification were done as show below



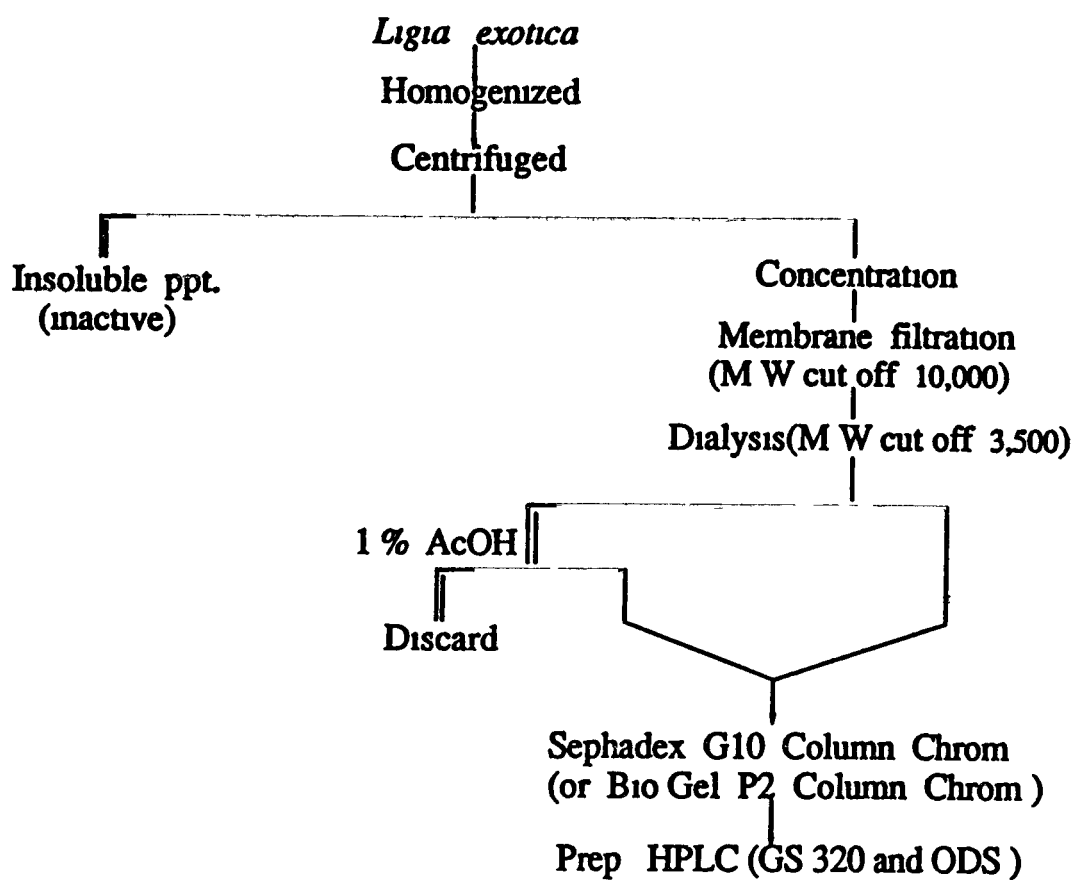
Doridosine

** Isolation and Purification of Doridosine



3 Separation and purification from *Ligia exotica*

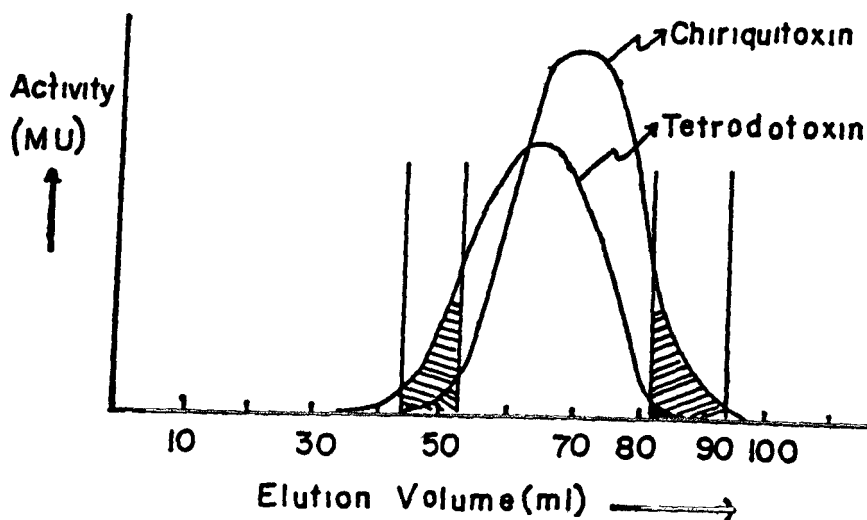
Biologically active compounds (inhibitory effects on histamine release), amino acids, and nucleic acids were separated and identified



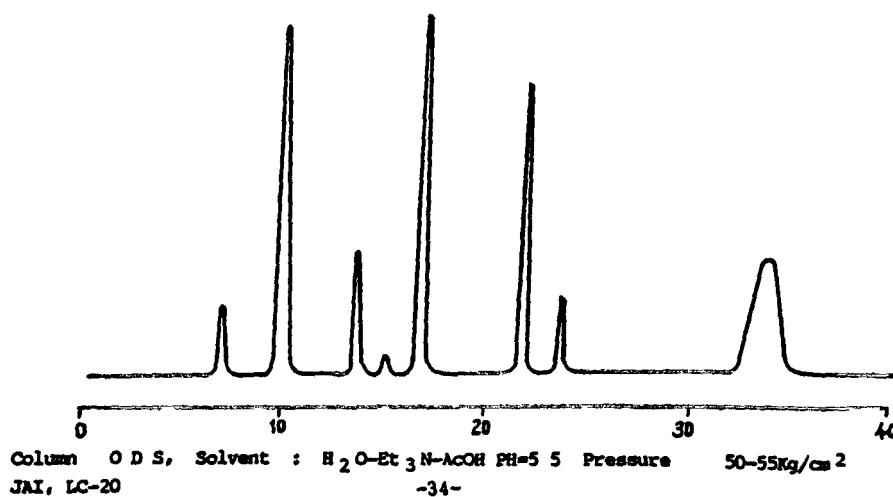
4 Separation and characterization of Tetrotoxin (TTX) and Chiriquitoxin (CHTX)

summary The structure of chiriquitoxin, a tetrodotoxin analog isolated from the Costa Rican frog Atelopus chiriquiensis, was elucidated on the basis of NMR data. In the structure 11-CH₂OH of tetrodotoxin was replaced by a CH(OH)CH(NH₂)COOH group

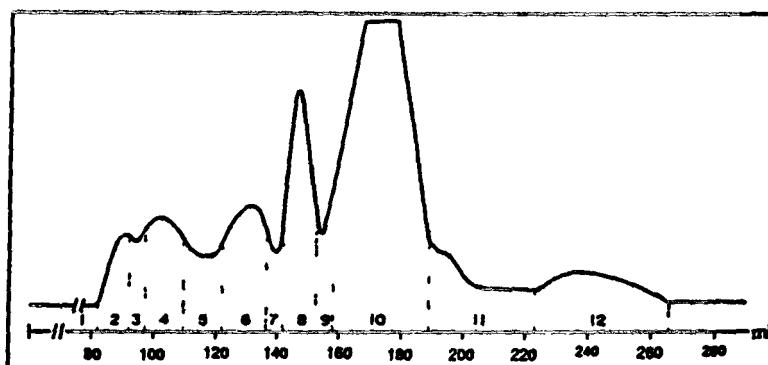
Bio-Gel P-2 Filtration By LC



** Purifies from crude TTX and CHTX



Sephadex G 10 Column Chromatography



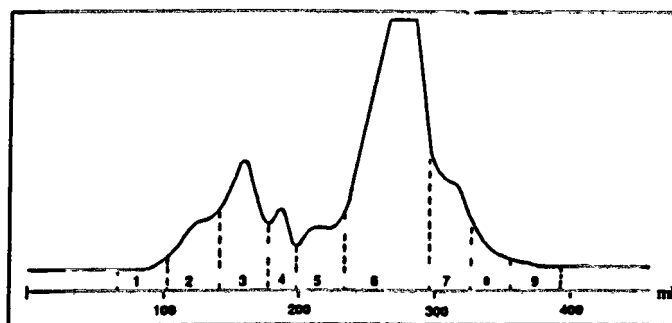
Column Sephadex G 10 (1.9 X 110 cm)
Detector UV 254
Eluent H₂O
Flow rate 20 ml/hr
Sample 2g/3ml H₂O

PREPARATIVE HPLC SEPARATION

Column JAI GS 320 (20X500 mm)
Detector UV 254 and RI
Eluent MeOH H₂O = 70 30
Flow rate 3 ml/min

Sample No	Isolated Compounds
6	Betaine, Ala, Val, Pro Ile, Leu, Gly, Unknown 1, 2
8	Phe, Uracine, Unknown 3, 4
10	Inosine, Unknown 5, 6
12	Trp-1, Guanosine, Unknown 7 2-Deoxyguanosine
ppt	Tyrosine

LOW PRESSURE LC



Column Bio Gel P2 (20X300 mm)
 Detector UV 254
 Eluent Et₃N-AcOH-H₂O pH 6.2
 Flow rate 3 ml/min

Fraction NO	1	2	3	4	5	6	7	8	9
Amounts (mg)	-	16	49	59	7	19	7	4	9

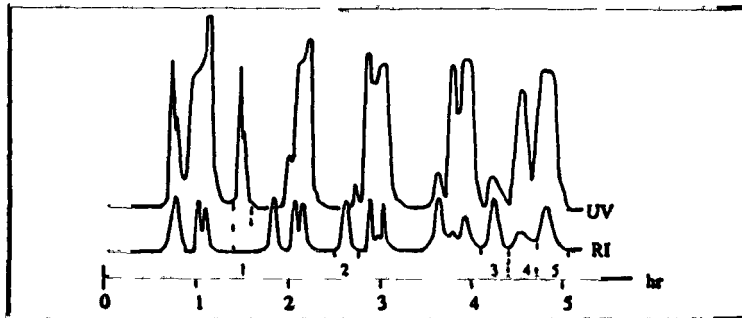
BIOLOGICAL ACTIVITY

Inhibitory effects of *Ligula Exonca* fractions on Histamine release
from Rat Peritoneal Mast Cells induced by compound 48/80

Treatment	Concentration	Histamine release (%)	Inhibition (%)
Control		68.3	
Blank		3.6	
1		26.0	6.4
2	0.5	19.4	75.6
3	0.5	12.6	86.1
4	0.5	12.5	86.2
5		42.9	39.3
6	0.5	12.4	86.4
7	0.5	18.1	77.6
8	0.5	60.3	12.3
9	0.5	32.5	55.2

*mg/ml Concentration of compound 48/80 (5×10^{-7} g/ml)

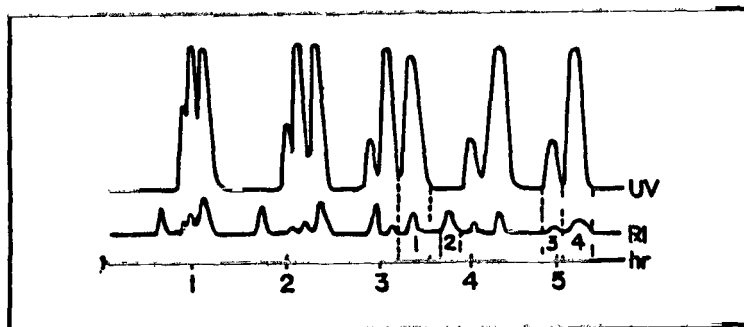
RECYCLE PREPARATIVE HPLC



Sample Sephadex G 10 Column Fraction 10

Fraction No	Isolated compounds
1	Unknown 7
2	Ile, Leu
3	Phenylalanine
4	Unknown 8
5	Inosine

RECYCLE PREPARATIVE HPLC



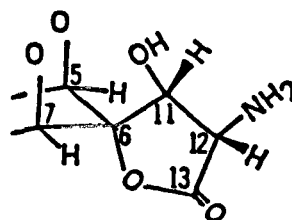
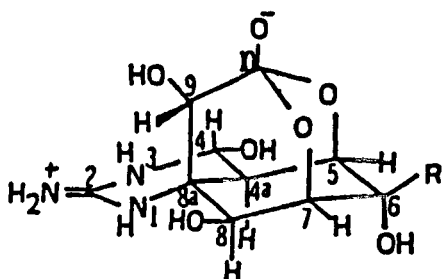
Sample Sephadex G 10 Column Fraction 12

Fraction No	Isolated compounds
1	Trp-1
2	Unknown 7
3	2-Deoxyguanosine
4	Guanosine

NMR spectral data of CHTX and CHTX-13,6-lactone

CHTX (1)*			CHTX-13,6-lactone (4)**		
	C	H		C	H
2	156.6			156.7	
4	75.2	5 51 (d 9 4)		75.3	5 53 (d 8 5)
4a	40.5	2 31 (d 9 4)		41.7	2 16 (d 8 5)
5	73.5	4 40 (br s)		70.6	4 71 (br s)
6	72.1			84.3	
7	81.1	4 39 (br t)		77.7	4 30 (br t)
8	72.7	4 40 (br s)		72.8	4 19 (d 1 5)
8a	59.3			59.5	
9	70.9	4 00 (s)		70.7	4 03 (s)
10	111.0			111.1	
11	70.3	4 90 (d 1 8)		69.7	5 20 (d 5 5)
12	58.1	4 27 (d 1 8)		-	4 85 (d 5 5)
13	174.1			173.0	

¹³C NMR *75.5 MHz (GN-300), **100 MHz (JEOL GSX-400)
¹³CD₃COOD as 22.4 ppm ¹H NMR *300 MHz (GN-300),
**400 MHz (JEOL GSX-400), CHD₂COOD as 2.06 ppm
Solvent * 4%CD₃COOD/D₂O, ** 1%TFA, 4%CD₃COOD/D₂O (45°C)
- Unassignable carbon due to an exchange of H-12 with D
(¹³C NMR spectrum of 4 was measured after keeping 1 in
1%TFA, 4%CD₃COOD/D₂O for one month at 5°C)



	R
1 CHTX	11 CH(OH)CH(NH ₂)COOH 12 R 13 S
2 TTX	11 CH ₂ OH
3 11-norTTX-6,6-diol	OH

4 CHTX-13,6-lactone

References

- 1 Y H Kim, F A Fuhrman, G J Fuhrman, L A Pavelka, and H S Mosher, *Science*, **207**, 194 (1980)
- 2 R J Quinn, R P Gregson, A F Cook, and R T Bartlett, *Tetrahedron Lett.*, **567** (1980)
- 3 R P Gregson, R J Quinn, and A F Cook, German Patent No 2,833,887 issued Feb 2, 1979, *Chem Abst*, **91**, 39792 (1979)
- 4 K Grozinger, K P Freter, P Farina, and A Gladczuk, *Eur, J Med Chem Ther*, **18**, 221 (1983)
- 5 Unpublished data
- 6 Unpublished data
- 7 G R McDonald and J W Nybakken, *The Veliger*, **21**, 110 (1979)
- 8 S A Bloom, *The Veliger*, **13**, 289 (1976)
- 9 J Baird-Lambert, J F Marwood, L P Davies, and K M Taylor, *Life Sciences*, **26**, 1069-1077 (1980)
- 10 L P Davies, K M Taylor, R P Gregson, and R J Quinn, *Life Sciences*, **26**, 1079-1088 (1980)
- 11 Y H Kim, G H Brown, H S Mosher, F A Fuhrman, *science*, **189**, 151 (1975)
- 12 L A Pavelka, Y H Kim, H S Mosher, *Toxicon*, **15**, 135 (1976)
- 13 R D Macfarlane, D F Torgerson, *science*, **191**, 920 (1976)
- 14 (a) K. Tsuda, S Ikuma, M Kawamura, R Tachikawa, K Sakai, C Tamura, O Amakasu, *Chem Pharm Bull*, **12**, 1357 (1964) (b) R B Woodward, *Pure Appl Chem*, **9**, 49 (1964) (c) T Goto, Y Kishi, S Takahashi, Y Hirata, *Tetrahedron*, **21**, 2059 (1965)
- 15 H S Mosher, F A Fuhrman, H D Buchwald, H G Fischer, *science*, **114**, 1100 (1964)
- 16 C Y Kao, *Pharm Rev*, **18**, 997 (1966)
- 17 (a) T Yasumoto, M Yotsu, M Murata, H Naoki, *J Am Chem Soc*, **110**, 2344 (1988) (b) M Nakamura, T Yasumoto, *Toxicon*, **23**, 271 (1985) (c) A Endo, S S Khora, M Murata, H Naoki, T Yasumoto, *Tetrahedron Lett*, **29**, 4127 (1988) (d) S S Khora, T Yasumoto, *ibid*, **30**, 4393 (1989)
- 18 C Y Kao, P N Yeoh, M D Goldfinger, F A Fuhrman, H S Mosher, *J Pharmacol Exp Therap*, **217**, 416 (1981)
- 19 C Y Kao, T Yasumoto, *Toxicon*, **23**, 725 (1985)